**SAFETY Meeting Minutes**
UAMS IBC

## MEETING TIME RECORDS

**Meeting start time:** 1/9/2026 12:02 PM
**Meeting end time:** 12:16 PM
**Meeting type:** Virtual

| Name of Regular/Alternate Member | Status (Member or Alternate) | Present by Teleconference? |
|---|---|---|
| Ha-Neui Kim | Member | No |
| Matthew Jorgenson | Member | Yes |
| Robert Hunter | Member | No – voted by e-mail |
| Kimberly Murphy | Member | Yes |
| Lindsey Clark | Member | Yes |
| James Douglas | Member | Yes |
| Amanda Holloway | Ex Officio | No |
| James Bishop | Member | No |
| Youssef Aachoui | Member | Yes |
| Jia Liu | Member | Yes |
| Yuet-Kin Leung | Member | Yes |
| Melaney Gee | Member | Yes |
| Mark Manzano | Member | Yes |
| Christine Simecka Morgan | Member | No |
| Antino Allen | Member | Yes |
| KyoungHyun Kim | Member | No |
| James Townsend | Ex Officio | Yes |
| Shengyu Mu | Member | No |
| Kikumi Ono-Moore | Ex Officio | Yes |
| Zhiqiang Qin | Member | Yes |

## QUORUM INFORMATION

**Number of SAFETY members on the roster:** 17
**Number required for quorum:** 9
**Quorum:** Present

All members present via teleconference received all pertinent material before the meeting and were able to actively and equally participate in all discussions.

| ATTENDANCE STATUS AND VOTING KEY | |
|---|---|
| ABSTAIN: | Present for the vote but not voting "For" or "Against." |
| ABSENT: | Absent for discussion and voting for reasons other than a conflict of interest. |
| RECUSED: | Absent from the meeting during discussion and voting because of a conflict of interest. |
| SUBSTITUTION: | When regular members and their alternate(s) are listed in the ATTENDANCE table above and an alternate member serves as a substitute for the regular member this identifies the name of the alternate to indicate which individual is serving as the voting member for this vote. May be deleted if there are no substitutions. |

| GUEST NAMES |
|---|
| N/A |
| |

**Previous Meeting minutes approved:** Yes

## REVIEW OF SUBMISSIONS

The review and discussion of the protocols listed below included the following elements: the agents involved and their characteristics; types of manipulations planned; the source(s) and nature of the nucleic acid sequences; the host organism(s) and vector(s) to be utilized; whether expression of a foreign gene is intended and, if so, the specific protein(s) to be produced; the containment conditions to be applied, including biosafety level and any special provisions; and the relevant sections of the NIH Guidelines.

All IBC members present were reminded to identify any conflicts of interest as each registration was reviewed.

For each protocol reviewed, it was confirmed that the Principal Investigator (PI) and laboratory personnel have received appropriate training in the safe conduct of research.

**De Novo Review**

**1. Review of SPROTO202500000076**

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| Title: | Mechanisms of Erythroid Remission in Diamond Blackfan Anemia (DBA) (BP254) |
| Investigator: | Jason Farrar |
| Submission ID: | SPROTO202500000076 |
| Description: | This research lab and the projects herein investigate and |

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| | support childhood blood and cancer-related disorders study. Current projects include the investigation of genetic determinants of Diamond Blackfan anemia as well as maintenance of a blood and tissue repository. Agents of biosafety concern include: human blood, human tissue, human cell lines, lentivirus, rDNA, and E. coli.<br><br>Human tissues and blood are collected in a clinical environment and then either shipped to the lab by IATA compliant measures or transported on-campus in leak-proof containers as outlined below. Frozen specimens obtained from the pathology lab are transported on campus on dry-ice also in IATA compliant shipping containers.<br><br>Specimens are processed in the tissue culture laboratory in certified Class II BSC or directly stored in -80 or liquid nitrogen containers. Where needed, analysis and sorting of human cells or cell lines by flow cytometry is performed in a dedicated BSL-2-certified cabinet validated for the sorting device. Laboratory safety, PPE, is required to protect the wearer from exposure to hazardous materials.<br><br>Cell cultures are centrifuged with sealed vessels. Loading/un-loading of primary sealed containers are performed into rotor buckets inside the BSC using safety caps. The transportation of samples is triple packed; carried in a container that is leak proof which includes absorbent material to contain liquid if the primary container breaks. The outer container is labeled with a biohazard label.<br><br>E coli is used for routine cloning of human gene products and propagation of plasmids and lentiviral vectors. There is no manipulation or alteration of bacterial or viral virulence factors or toxins; these studies are all directed at biochemical characterization and functional testing in vivo cultured human cell lines or primary cells to determine the role of human genes involved in erythropoiesis and formation of cancer.<br><br>Lentiviral transduction systems are used to alter cell lines and primary cells in short term culture to assess the effect of key erythroid or other genes implicated in formation of cancer and in differential erythroid responses during bone marrow failure. We may employ small interfering RNA, Cas9 based genomic editing, epigenomic editing and forced expression of identified genes in these short term culture systems of human cell lines or primary cell cultures. There is no transfer of human cells or |

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| | rDNA to animal models planned. |
| Agent Containment: | Biological Containment Levels:<br>• Cerebrospinal Fluid (CSF): BSL-2<br>• Human Tumor Tissue: BSL-2<br>• Buccal Swab: BSL-2<br>• Human Blood: BSL-2<br>• Human Serum: BSL-2<br>• Urine: BSL-2<br>• Lentivirus: BSL-2<br>• E. coli: BSL-2<br>• K-562 Cell Line (Human Lymphoblasts): BSL-2<br>• Human Red Blood Cells: BSL-2<br>• HEK293T Human Cell Line: BSL-2 |
| Applicable NIH Guidelines: | • Section III-D-2-a<br>• Section III-D-3-a |

a. **Determination:** Modifications Required

b. **Required modifications:**
Please respond to comments and make any necessary modifications.

c. **Votes:**

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| **For:** | 11 |
| **Against:** | 0 |
| **Recused:** | 0 |
| **Absent:** | 6 |
| **Abstained:** | 0 |

**De Novo Review**

**2. Review of SPROTO202500000085**

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|---|---|
| Title: | Infection and lipoproteins |
| Investigator: | Ryan Allen |
| Submission ID: | SPROTO202500000085 |
| Description: | This research will seek to understand how lipoproteins contribute to the host-response to infection in mice. We will combine genetic and environmental manipulation of lipoprotein metabolism with in vivo models of infection to study how lipoproteins contribute to dysfunctional host responses that phenocopy sepsis in humans. Some experiments will utilize adeno-associated viral vectors (AAV8-CETPxAPOE3*Leiden or AAV8-PCSK9) to humanize the mouse lipoprotein profile. We will use intra-tracheal inoculation with Klebsiella pneumoniae or staphylococcus |

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| | aureus to induce bacterial pneumonia in mice and study how changes in lipoprotein metabolism influence the host inflammatory response. Alternatively, we will introduce S. aureus directly to bone (osteomyelitis) or to the ventricular cavity of mice (endocarditis secondary to bacteremia).<br><br>All bacteria listed above will be cultured in plastic dishes within a sealed incubator and handled within a BSL-2 certified biosafety cabinet. When centrifugation is necessary, cells will be transferred to conical tubes, sealed with parafilm and transferred to a desktop centrifuge in the lab. AAV will be purchased from a commercial vendor and stored at -80C. When necessary, AAV will be thawed and aliquoted in a BSL-2 biosafety cabinet. Animal tissues and blood will be harvested in designated clean areas covered with absorptive bench pads. Fresh tissue may be homogenized to single cell suspensions and serially diluted prior to plating on selective media to quantify pathogen colony forming units. Animal tissues may be flash frozen in liquid nitrogen and when necessary, pulverized prior to RNA, protein or lipid extraction. Frozen plasma may be thawed, sterile filtered, and injected to an AKTA Pure fast-protein liquid chromatography system (FPLC) and fractionated to separate lipoprotein pools. |
| Agent Containment: | Biological Containment Levels:<br>• Bronchoalveolar lavage: BSL-2<br>• Animal Tissue: BSL-2<br>• Animal Serum: BSL-2<br>• Adenoassociated virus AAV: BSL-2<br>• Adenoassociated virus AAV: BSL-2<br>• Staphylococcus aureus: BSL-2<br>• Klebsiella: BSL-2<br>• Staphylococcus aureus: BSL-2<br>• Staphylococcus aureus: BSL-2 |
| Applicable NIH Guidelines: | • Section III-D-4<br>• Section III-D |

a. **Determination:** Modifications Required

b. **Required modifications:**
Please respond to comments and make any necessary modifications.

c. **Votes:**

| | |
|---|---|
| **For:** | 11 |
| **Against:** | 0 |
| **Recused:** | 0 |
| **Absent:** | 6 |
| **Abstained:** | 0 |

## REVIEW OF OTHER AGENDA ITEMS

- Administrative approvals were acknowledged.
- Potential safety events were reported for SPROTO202400000042 and SPROTO202500000013 as it related to ABLS-2 procedures not being followed. Animals may not have had BSL-2 cage cards and were not housed in appropriate location. No exposures reported. BSOs will investigate and report back to committee at next month's meeting.
- No inspection findings to report.
- Dr. Liu is still working on community member recruitment.
- No other new business was discussed